Cerebral venous thrombosis and portal vein thrombosis: a retrospective cohort study of 537,913 COVID-19 cases

Maxime Taquet, 1,2,* Masud Husain, 3,4 John R Geddes, 1,2 Sierra Luciano, 5 Paul J Harrison 1,2

¹Department of Psychiatry, University of Oxford, Oxford, UK

²Oxford Health NHS Foundation Trust, Oxford, UK

³Nuffield Department of Clinical Neurosciences, University of Oxford, Oxford, UK

⁴Oxford University Hospitals NHS Foundation Trust, Oxford, UK

⁵TriNetX, Cambridge MA, USA

*Corresponding author: University Department of Psychiatry, Warneford Hospital, Oxford OX3 7JX, UK. maxime.taquet@mesdsci.ox.ac.uk

Maxime Taquet: Academic Clinical Fellow, University of Oxford

Masud Husain: Professor of Neurology and Cognitive Neuroscience, University of Oxford

John R Geddes: Professor of Epidemiological Psychiatry, University of Oxford Sierra Luciano: Content Manager, Clinical Sciences and Operations, TriNetX, USA

Paul J Harrison: Professor of Psychiatry, University of Oxford

Abstract

Objectives: To estimate the absolute risk of cerebral venous thrombosis (CVT) and portal vein thrombosis (PVT) in the two weeks following a diagnosis of COVID-19, and to assess the relative risks (RR) compared to influenza or the administration of an mRNA vaccine against COVID-19.

Design: Retrospective cohort study based on an electronic health records network

Setting: Linked records between primary and secondary care centres within 59 healthcare organisations, primarily in the USA

Participants: All patients with a confirmed diagnosis of COVID-19 between January 20, 2020 and March 25, 2021 were included (N=537,913, mean [SD] age: 46.2 [21.4] years; 54.9% females). Cohorts (matched for age, sex, and race) of participants diagnosed with influenza (N=392,424) or receiving the BNT162b2 or mRNA-1273 vaccine (N=366,869) were used for comparison.

Main outcome measures: Diagnosis of CVT (ICD-10 code I67.6) or PVT (ICD-10 code I81) within 2 weeks after a diagnosis of COVID-19.

Results: The incidence of CVT after COVID-19 diagnosis was 42.8 per million people (95% CI 28.5–64.2) including 35.3 per million (95% CI 22.6–55.2) first diagnoses. This was significantly higher than the CVT incidence in a matched cohort of patients with influenza (RR=3.83, 95% CI 1.56–9.41, P<0.001) and people who received an mRNA vaccine (RR=6.67, 95% CI 1.98–22.43, P<0.001). The incidence of PVT after COVID-19 diagnosis was 392.3 per million people (95% CI 342.8–448.9) including 175.0 per million (95% CI 143.0–214.1) first diagnoses. This was significantly higher than the PVT incidence in a matched cohort of patients with influenza (RR=1.39, 95% CI 1.06–1.83, P=0.02) and people who received an mRNA vaccine (RR=7.40, 95% CI 4.87–11.24, P<0.001). Mortality after CVT and PVT was 17.4% and 19.9% respectively.

Conclusions: The incidence of CVT and PVT is significantly increased after COVID-19. The data highlight the risk of serious thrombotic events in COVID-19 and can help contextualize the risks and benefits of vaccination in this regard.

What is known

- A systematic review of cohort studies suggested an incidence of CVT among *hospitalised* patients with COVID-19 to be about 800 per million patients. There was evidence of selection, ascertainment, and reporting bias in all included studies.
- The incidence of CVT and PVT among both hospitalised and non-hospitalised patients with COVID-19 is unknown.
- It is unknown if COVID-19 increases the risk of CVT and PVT.

What this study adds

Our study estimates that the absolute risk of CVT and PVT are respectively 42.8 and 392.3 per million patients (both hospitalised and non-hospitalised) in the 2 weeks after a diagnosis of COVID-19. COVID-19 increases the risk of CVT and PVT compared to patients diagnosed with influenza, and to people who have received a COVID-19 mRNA vaccine.

There are concerns about a possible association between vaccines against SARS-CoV-2 and cerebral venous thrombosis (CVT, also called cerebral venous sinus thrombosis [1]). The concern has focused primarily on ChAdOx1 nCoV-19 ("Oxford-AstraZeneca") vaccine, and more recently the Ad26.COV2.S ("Janssen") vaccine. Emerging data suggest that the association reflects a 'vaccine-induced thrombotic thrombocytopaenia' (VITT) [2,3]. Governments and regulatory authorities have reacted by restricting the use of the two vaccines in different subgroups of the population, based on a risk-benefit analysis. Yet one key component of the risk-benefit calculation is currently unknown: the absolute risk of CVT following a diagnosis of COVID-19. To date there are only a few case series of CVT post-COVID-19, and a few cohort studies limited to hospitalised patients and with evidence of selection, ascertainment, and reporting bias [4].

Here, using an electronic health records network primarily based in the USA, we estimated the incidence of CVT occurring in confirmed COVID-19 cases (both hospitalised and non-hospitalised) and compared this incidence to two other groups: people who received a COVID-19 mRNA vaccine, and a cohort of patients with influenza. We could not make a direct comparison with rates after the ChAdOx1 nCoV-19 ("Oxford-AstraZeneca") vaccine because this has not been used in the USA. We also examined the rate of portal vein thrombosis (PVT), another diagnosis associated with thrombosis in the venous system and thought to occur in VITT [3].

Methods

Data

We used TriNetX Analytics, a federated electronic health records network recording anonymized data from 59 healthcare organizations, primarily in the USA, totalling 81 million patients. For details, see [5], and the supplement.

The process by which the data is de-identified is attested to through a formal determination by a qualified expert as defined in Section §164.514(b)(1) of the HIPAA Privacy Rule, so that ethical approval from an institutional review board is not needed.

Primary analysis

A cohort of all patients who had a confirmed diagnosis of COVID-19 (ICD-10 code U07.1) between January 20, 2020 and March 25, 2021 was defined for study. The absolute risk of a diagnosis of CVT (ICD-10 code I67.6) was calculated by identifying those patients in the cohort who had the diagnosis in the two weeks following their diagnosis of COVID-19. The absolute risk of PVT (ICD-10 code I81) was also calculated. For the whole COVID-19 cohort, and for cases with CVT or PVT following COVID-19, baseline characteristics are reported. We identified patients who had a reported high D-dimer (> 5mg/L), low fibrinogen (< 200 mg/dL), or thrombocytopenia within the 2 weeks after their COVID-19 diagnosis. We also assessed how many of them had died (and, if so, when) by the time of the analysis (April 21, 2021).

A causal link between COVID-19 and CVT/PVT cannot be established with a simple cohort study. However, a testable corollary of such a causal association is that the rate of new CVT/PVT diagnoses decreases with time from the index event. We tested this corollary by comparing the absolute risk within two weeks of diagnosis (Week 1 and 2) with the absolute

risk with the next two weeks (Week 3 and 4) and the two weeks thereafter (Week 5 and 6). For this part of the analysis, only patients diagnosed on or before February 28, 2021 were included to allow for sufficient follow-up.

Two control cohorts based on other index events were used for comparison: a diagnosis of influenza (ICD-10 codes J09-J11) between January 20, 2018 and March 25, 2021 (an earlier start date was used for this event to achieve a sufficiently large sample), and receipt of a first dose of the two vaccines administered to this predominantly US population: the BNT162b2 ('Pfizer-BioNTech') vaccine or the mRNA-1273 ('Moderna') vaccine before March 25, 2021. We excluded from these cohorts any patients who had a diagnosis of COVID-19 on or after January 20, 2020.

These two cohorts were then matched to the cohort of patients with COVID-19 for age (defined as a continuous variable), sex (defined as a categorical variable taking the value: female, male, or other), and race (defined as a categorical variable taking the value: White, Black or African American, Asian, American Indian or Alaska Native, Native Hawaiian or Other Pacific Islander, or Unknown) using propensity score matching (see details below). Using these matched cohorts, we calculated the relative risk (RR) of a CVT diagnosis, a PVT diagnosis, and a diagnosis of thrombocytopenia in the two weeks after COVID-19 diagnosis compared to the other index events (i.e. influenza or vaccination).

Secondary analyses

The analyses were repeated after broadening the diagnostic criteria for CVT to include I63.6 (cerebral infarction due to central thrombosis, non-pyogenic), G08 (intracranial and intraspinal phlebitis and thrombophlebitis), O22.5 (CVT in pregnancy) and O87.3 (CVT in the puerperium), in line with recent epidemiological studies that have taken this definition of CVT in other settings [6,7].

The analyses were also repeated after excluding those patients who had had a prior diagnosis of the event of interest (CVT or PVT).

Statistical analyses

Fisher's exact tests were used to compare characteristics (baseline and laboratory) and death rates between patients with COVID-19 who had a CVT (or PVT) compared to patients with COVID-19 who did not. Fisher's exact tests were also used to test the null hypothesis that the relative risks of CVT and PVT in the two weeks after COVID-19 vs. influenza and vs. mRNA vaccine were equal to 1. Confidence intervals for absolute risks were based on Wilson score intervals. Confidence intervals for relative risks were based on Wald confidence limits, with Agresti-Coull adjustment to improve coverage when the risks are small [8].

Propensity score matching was used to create cohorts with matched baseline characteristics, and carried out within the TriNetX network. Propensity score 1:1 matching used a greedy nearest neighbour matching approach with a caliper distance of 0.1 pooled standard deviations of the logit of the propensity score. Any characteristic with a standardized mean difference (SMD) between cohorts lower than 0.1 is considered well matched. [9]

Statistical significance was set at a 2-sided P value < 0.05. Analyses were performed using R version 3.6.3. This study follows the STROBE reporting guidelines (see Supplement for a

checklist). Further details about TriNetX, cohort definitions, and statistical analyses can be found in the supplement.

Patient and Public Involvement

Patients and public were not involved in this study.

Results

537,913 patients with a confirmed diagnosis of COVID-19 were included in this study (54.9% females, mean [SD] age 46.2 [21.4]; Table 1). Of these, 23 were diagnosed with a CVT in the two weeks following their diagnosis (absolute risk: 42.8 per million people, 95% CI 28.5–64.2, equivalent to an incidence of 111.5 per 100k person-years). The risk was significantly higher among patients with a history of cardiovascular diseases (Table 1), specifically arterial diseases, cerebral/precerebral artery stenosis/occlusion, and intracranial haemorrhage.

Among the 23 events, 7 were observed in patients under the age of 30, 4 between 30 and 39, 2 between 40 and 49, 3 between 50 and 59, 2 between 60 and 69, and 5 between 70 and 79. Four patients had also had a CVT diagnosed prior to their COVID-19 diagnosis, one between 4 and 8 weeks beforehand, and the other 3 more than 8 weeks prior. The incidence of CVT following COVID-19 significantly decreased with time from the index event (RR in 3rd and 4th week vs. first 2 weeks 0.24, 95% CI 0.098–0.59, P<0.001; RR in 5th and 6th week vs. first 2 weeks 0.12, 95% CI 0.036–0.40, P<0.001; Fig. 1).

The absolute risk of PVT in the two weeks following COVID was 392.3 per million people (95% CI 342.8–448.9), equivalent to 1022.7 per 100k person-years. Among the 211 affected patients, 117 had a PVT prior to their COVID-19 diagnosis. The incidence of PVT following COVID-19 significantly decreased with time from the index event (RR in 3rd and 4th week vs. first 2 weeks 0.19, 95% CI 0.14–0.27, P<0.001; RR in 5th and 6th week vs. first 2 weeks 0.12, 95% CI 0.080–0.18, P<0.001; Fig. 1).

Laboratory data were available for a subset of the COVID-19 patients (Table 2). Although the data do not cover most patients with a diagnosis of CVT, they suggest that patients with CVT after COVID-19 were significantly more likely to have elevated D-dimer level than patients with COVID-19 who did not have CVT, whereas patients with PVT after COVID-19 were significantly more likely to have low fibrinogen level and thrombocytopenia. The death rate among patients with CVT in the two weeks after COVID-19 was 17.4% (4 out of 23 patients, 95% CI 6.98–37.1%; Figure S1 in the supplement) and that among patients with PVT after COVID-19 was 19.9% (42 out of 211 patients, 95% CI 15.1–25.8%; Figure S1 in the supplement) and were significantly higher than among patients with COVID-19 who did not have those events (P=0.005 for CVT and P<0.001 for PVT).

The two-week risk of being diagnosed with a CVT was significantly higher in the cohort diagnosed with COVID-19 compared to a matched cohort diagnosed with influenza (N=392,424 in each cohort; RR=3.83, 95% CI 1.56–9.41, P<0.001; Fig. 2 and Table S1) or compared to a matched cohort receiving an mRNA vaccine (N=366,869 in each cohort; RR=6.67, 95% CI 1.98–22.43, P<0.001; Fig. 2 and Table S2). Similarly, the two-week risk of being diagnosed with a PVT was significantly higher in the cohort diagnosed with COVID-19 compared to a matched cohort diagnosed with influenza (RR=1.39, 95% CI 1.06–1.83, P=0.02; Fig. 2) or compared to a matched cohort receiving an mRNA vaccine (RR=7.40, 95% CI 4.87–

11.24, P<0.001; Fig. 2). A diagnosis of thrombocytopenia was also significantly more likely in the two weeks after a diagnosis of COVID-19 than after a diagnosis of influenza (RR=1.23, 95% CI 1.18–1.28, P<0.001) or after receiving an mRNA vaccine (RR=30.8, 95% CI 27.1–35.0, P<0.001).

When the definition of CVT in terms of ICD-10 codes was broadened, the incidence of CVT in the two weeks after COVID-19 was 148.7 per million people (95% CI 119.5–185.1), which was significantly higher than in the matched cohort of patients with influenza (RR=3.37, 95% CI 2.02–5.62; P<0.001) and the matched cohort of people receiving an mRNA vaccine (RR=5.90, 95% CI 3.02–11.53, P<0.001; Fig. 2). The majority of the extra cases came from the G08 diagnostic category.

Finally, we excluded patients who had also had a CVT or PVT prior to COVID-19. The incidence of CVT and PVT post-COVID-19 diagnosis were reduced accordingly (CVT: absolute risk 35.3 per million, 95% CI 22.6–55.2); PVT (absolute risk 175.0 per million, 95% CI 143.0–214.1), but all RRs were similar to those in the primary analysis (Fig. 2).

Discussion

In a large electronic health records network, the absolute incidence of CVT and PVT in the 14 days after COVID-19 diagnosis was 42.8 and 392.3 per million patients respectively. The incidence rapidly decreased in the following weeks, which is compatible with a causal link between COVID-19 and those thrombotic events. However, causation cannot be demonstrated with the current study and residual confounding (e.g. increased medical monitoring directly after COVID-19 vs. a few weeks later) might contribute to this observation.

The incidence of CVT and PVT after COVID-19 is substantially greater than in the matched control cohorts. The incidence of CVT after a diagnosis of COVID-19 is also substantially greater than the expected incidence in the general population in the USA, estimated to be between 0.53 and 0.77 per million people in any 2-week period [7] and the rate is significantly higher than the highest of these estimates (binomial test: P<0.001).

The incidence is also many-fold higher than the latest reported incidence of CVT following administration of the first dose of the ChAdOx1 nCoV-19 ('Oxford-AstraZeneca') vaccine (reported by the European Medicines Agency to be around 5 per million vaccinated people [10]) and the latest reported incidence of CVT following administration of the Ad26.COV2.S ("J&J") vaccine (reported by the Food and Drug Administration to be about 0.9 per million vaccinated people [11]).

The increased rate of CVT in COVID-19 is notable, being much more marked than the increased risks for other forms of stroke and cerebral haemorrhage [5]. The PVT data highlight that COVID-19 is associated with thrombotic events that are not limited to the cerebral vasculature.

Importantly, the present study cannot be used to draw conclusions on the relative risk of developing a CVT or PVT after receiving an mRNA vaccine compared to the baseline incidence or compared to other vaccines. Far larger samples are needed (such as those used by the EMA and the FDA pharmacovigilance studies) because the events have so far been found to be extremely rare. The observed incidence of CVT in the matched cohort of people who

received an mRNA vaccine is compatible with even the lowest estimate of the baseline rate in the USA of 0.53 per million people in any 2-week period (binomial test: P=0.18 [7]).

The main strengths of this study are its large sample size and the use of matched cohorts. However, the study also has several limitations and results should be interpreted with caution. First, while cohorts were matched for age, sex, and race, they were not matched for comorbidities so that the latter might be contributing to the association between COVID-19 and subsequent CVT/PVT. Second, we have no information about diagnostic accuracy or completeness of records, though this is likely to be less of an issue for CVT or PVT compared to many diagnoses. Third, some cases of COVID-19, especially those early in the pandemic, are undiagnosed, and we cannot generalise our risk estimates to this population. Similarly, COVID-19 vaccines are also being delivered by sites which are not part of an HCO, and many of these may not be coded in the electronic health record, and so the relative risk estimates may not apply to them. Fourth, the absence of key haematological laboratory data from many patients limits our ability to comment on whether the mechanism of CVT after COVID-19 is likely to be similar or different from that observed after ChAdOx1 nCoV-19 or Ad26.COV2.S. In particular, we do not have information regarding anti-platelet factor 4 (PF4) antibodies that have been associated with VITT [2,3].

In summary, COVID-19 is associated with a markedly increased incidence of CVT compared to patients with influenza, people who have received BNT162b2 or mRNA-1273 vaccines and compared to the best estimates of the general population incidence. The risk with COVID-19 also appears greater than with ChAdOx1 nCoV-19 and Ad26.COV2.S vaccines, although as noted this conclusion is indirect and tentative. The rarity of CVT in all populations means that larger sample sizes are required to confirm the results, and complementary study designs are needed to aid interpretation. Nevertheless, the current data highlight the risk of serious thrombotic events in COVID-19, and can help contextualize and inform debate about the risk-benefit ratio for current COVID-19 vaccines.

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Transparency declaration: The corresponding author is the guarantor and affirms that this manuscript is an honest, accurate, and transparent account of the study being reported; that no important aspects of the study have been omitted; and that any discrepancies from the study as planned have been explained.

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Declaration of interests: All authors have completed the Unified Competing Interest form (available on request from the corresponding author) and declare: no support from any organisation for the submitted work [or describe if any]; no financial relationships with any organisations that might have an interest in the submitted work in the previous three years [or describe if any], no other relationships or activities that could appear to have influenced the

submitted work [or describe if any]." Please note: The corresponding author must collect Unified Competing Interest forms from all authors and summarise their declarations as above within the manuscript.SL is an employee of TriNetX.

Data sharing: The TriNetX system returned the results of the analyses as .csv files, which were downloaded and archived. Data presented will be freely accessible at: [the URL will be added upon publication]. In addition, TriNetX will grant access to researchers if they have a specific concern (through a third-party agreement option).

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Dissemination declaration: We plan to disseminate the results to the public and patients and family members of patients with COVID-19.

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Table 1 – Baseline characteristics of the whole COVID-19 cohort and the groups who received a diagnosis of CVT or PVT in the two weeks after COVID-19 diagnosis. The P-values from Fisher exact test (or t-test for age) for CVT and PVT groups compared to the rest of the COVID-19 cohort are shown.

| | All patients with COVID-19 | Patients with COVID-19 and CVT | | Patients with COVID-19 and PVT | |
|---------------------------------------|----------------------------|--------------------------------|---------|--------------------------------|---------|
| | n (%) mean (SD) | n (%) mean (SD) | P | n (%) mean (SD) | P |
| Sample size, n | 537913 (100.0) | 23 (100.0) | - | 211 (100.0) | - |
| Age, mean (SD), y | 46.2 (21.4) | 46.5 (21.5) | 0.95 | 57.2 (14.6) | < 0.001 |
| Sex, n (%) | | | | | |
| Female | 295220 (54.9) | 16 (69.6) | 0.21 | 94 (44.5) | 0.0029 |
| Male | 242439 (45.1) | 7 (30.4) | 0.21 | 117 (55.5) | 0.0028 |
| Race, n (%) | | | | | |
| White | 326258 (60.7) | 18 (78.3) | 0.091 | 145 (68.7) | 0.017 |
| Black | 93712 (17.4) | 4 (17.4) | 1 | 34 (16.1) | 0.72 |
| Asian | 14498 (2.7) | 2 (8.7) | 0.13 | 3 (1.4) | 0.39 |
| Other | 3909 (0.7) | 0 (0.0) | 1 | 1 (0.5) | 1 |
| Unknown | 99536 (18.5) | 2 (8.7) | 0.29 | 28 (13.3) | 0.051 |
| Comorbidities at baseline, n (%) | | | | | |
| Obesity | 92903 (17.3) | 4 (17.4) | 1 | 57 (27.0) | < 0.001 |
| Hypertension | 153305 (28.5) | 8 (34.8) | 0.49 | 115 (54.5) | < 0.001 |
| CKD | 35582 (6.6) | 2 (8.7) | 0.66 | 51 (24.2) | < 0.001 |
| Ischemic heart diseases | 48767 (9.1) | 5 (21.7) | 0.052 | 42 (19.9) | < 0.001 |
| Cardiac failure | 29536 (5.5) | 2 (8.7) | 0.36 | 26 (12.3) | < 0.001 |
| Arterial diseases | 38606 (7.2) | 7 (30.4) | < 0.001 | 44 (20.9) | < 0.001 |
| Venous diseases | 33222 (6.2) | 4 (17.4) | 0.05 | 171 (81.0) | < 0.001 |
| Cerebral/Pre-cerebral artery stenosis | 20817 (3.9) | 6 (26.1) | < 0.001 | 21 (10.0) | < 0.001 |
| Intracranial haemorrhage | 4056 (0.8) | 5 (21.7) | < 0.001 | 9 (4.3) | < 0.001 |
| Dementia | 11872 (2.2) | 1 (4.3) | 0.4 | 3 (1.4) | 0.64 |
| Chronic lower resp. disease | 90617 (16.8) | 6 (26.1) | 0.26 | 69 (32.7) | < 0.001 |
| Connective tissue disorders | 9671 (1.8) | 1 (4.3) | 0.34 | 15 (7.1) | < 0.001 |
| Liver disease | 32972 (6.1) | 2 (8.7) | 0.65 | 148 (70.1) | < 0.001 |
| Diabetes mellitus | 77038 (14.3) | 6 (26.1) | 0.13 | 73 (34.6) | < 0.001 |
| Malignancy | 40636 (7.6) | 3 (13.0) | 0.25 | 100 (47.4) | < 0.001 |
| Past CVT | 90 (0.02) | 4 (17.4) | < 0.001 | 0 (0.0) | 1 |
| Past PVT | 676 (0.1) | 0 (0.0) | 1 | 117 (55.5) | < 0.001 |

Table 2 – Laboratory characteristics of the patients in each group. P values are from Fisher's exact test, comparing the CVT and PVT groups to the rest of the COVID-19 cohort.

| | All patients with COVID-19 | Patients with COVID-19 and CVT | | Patients with COVID-19 and PVT | |
|--|----------------------------|--------------------------------|-------|--------------------------------|--------|
| | n (%) | n (%) | P | n (%) | P |
| D-dimer > 5 mg/L n/n with measurement (%) | 2047/69313 (3.0) | 2/6 (33.3) | 0.012 | 4/66 (6.1) | 0.14 |
| Fibrinogen < 200 mg/dL n/n with measurement (%) | 1167/19602 (6.0) | 1/6 (16.7) | 0.31 | 22/46 (47.8) | <0.001 |
| Thrombocytopenia (ICD-10 codes D69.49, D69.59, D69.6) | 8840 (1.8) | 0 (0.0) | 1 | 59 (28.0) | <0.001 |
| Death | 16619 (3.1) | 4 (17.4) | 0.005 | 42 (19.9) | <0.001 |

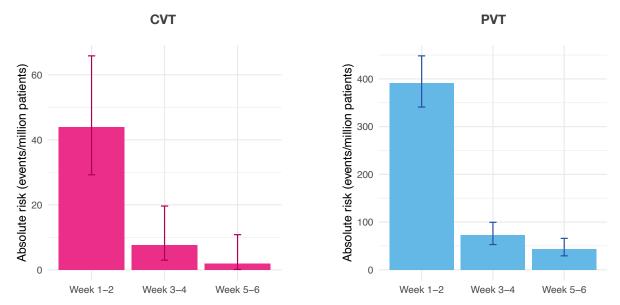


Figure 1 – Incidence of CVT and PVT per million patients as a function of the time since diagnosis of COVID-19, from the first 2 weeks post diagnosis to the 5th and 6th week post diagnosis.

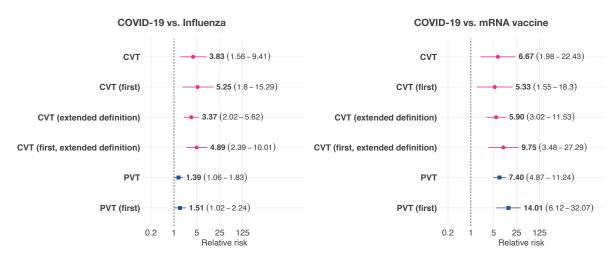


Figure 2 – Relative risk of CVT and PVT after a diagnosis of COVID-19 compared to matched cohorts of people with a diagnosis of influenza (left) or receiving an mRNA vaccine (right). Horizontal lines and numbers in brackets represent the 95% confidence intervals.

Supplement

Cerebral venous thrombosis and portal vein thrombosis: a retrospective cohort study of 537,913 COVID-19 cases

Maxime Taquet, 1,2,* Masud Husain, 3,4 John R Geddes, 1,2 Sierra Luciano, 5 Paul J Harrison 1,2

¹Department of Psychiatry, University of Oxford

²Oxford Health NHS Foundation Trust, Oxford, UK

³Nuffield Department of Clinical Neurosciences, University of Oxford, UK

⁴Oxford University Hospitals NHS Foundation Trust, Oxford, UK

⁵TriNetX, Cambridge MA, USA

sponding author: University Department of Psychiatry, Warneford Hospital, Oxford

*Corresponding author: University Department of Psychiatry, Warneford Hospital, Oxford OX3 7JX, UK. maxime.taquet@medsci.ox.ac.uk

Supplementary methods

TriNetX network

This section provides a version of our previous description of the network. [1]

Legal and ethical status

TriNetX's Analytics network is compliant with the Health Insurance Portability and Accountability Act (HIPAA), the US federal law which protects the privacy and security of healthcare data. TriNetX is certified to the ISO 27001:2013 standard and maintains an Information Security Management System (ISMS) to ensure the protection of the healthcare data it has access to and to meet the requirements of the HIPAA Security Rule. Any data displayed on the TriNetX Platform in aggregate form, or any patient level data provided in a data set generated by the TriNetX Platform, only contains de-identified data as per the de-identification standard defined in Section §164.514(a) of the HIPAA Privacy Rule. The process by which the data is de-identified is attested to through a formal determination by a qualified expert as defined in Section §164.514(b)(1) of the HIPAA Privacy Rule. This formal determination by a qualified expert, refreshed in December 2020, supersedes the need for TriNetX's previous waiver from the Western Institutional Review Board (IRB). The network contains data that are provided by participating Health Care Organizations (HCOs), each of which represents and warrants that it has all necessary rights, consents, approvals and authority to provide the data to TriNetX under a Business Associate Agreement (BAA), so long as their name remains anonymous as a data source and their data are utilized for research purposes. The data shared through the TriNetX Platform are attenuated to ensure that they do not include sufficient information to facilitate the determination of which HCO contributed which specific information about a patient.

Acquisition of data, quality control, and other procedures

The data are stored onboard a TriNetX appliance – a physical server residing at the institution's data centre or a virtual hosted appliance. The TriNetX platform is a fleet of these appliances connected into a federated network able to broadcast queries to each appliance. Results are subsequently collected and aggregated.

Once the data are sent to the network, they are mapped to a standard and controlled set of clinical terminologies and undergo a data quality assessment including 'data cleaning' that rejects records which do not meet the TriNetX quality standards. HIPAA compliance of the clinical patient data is achieved using de-identification. Different data modalities are available in the network. They include demographics (coded to HL7 version 3 administrative standards), diagnoses (represented by ICD-10-CM codes), procedures (coded in ICD-10-PCS or CPT), and measurements (coded to LOINC). While extensive information is provided about patients' diagnoses and procedures, other variables (such as socioeconomic and lifetime factors) are not comprehensively represented.

The data from a typical HCO generally go back around 7 years, with some going back 13 years. The data are continuously updated. HCOs update their data at various times, with most refreshing every 1, 2, or 4 weeks.

The data come primarily (>93%) from HCOs in the USA, with the remainder coming from India, Australia, Malaysia, Taiwan, Spain, UK, and Bulgaria. Only 1.8% of patients with COVID-19 are contributed from HCOs outside the USA. As noted above, to comply with legal frameworks and ethical guidelines guarding against data re-identification, the identity of participating HCOs and their individual contribution to each dataset are not disclosed to researchers.

Data quality assessment followed a standardised strategy wherein the data are reviewed for conformance (adherence to specified standards and formats), completeness (quantifying data presence or absence) and plausibility (believability of the data from a clinical perspective). There are pre-defined metrics for each of the above assessment categories. Results for these metrics are visualised and reviewed for each new site that joins the network as well as on an ongoing basis. Any identified issue is communicated to the data provider and resolved before continuing data collection.

The basic formatting of contributed data is also checked (e.g. to ensure that dates are properly represented). Records are checked against a list of required fields (e.g., patient identifier) and rejects those records for which the required information is missing. Referential integrity checking is done to ensure that data spanning multiple database tables can be successfully joined together. As the data are refreshed, changes in volume of data over time is monitored to ensure data validity. At least one non-demographic fact for each patient is required for them to be counted in the dataset. Patient records with only demographics information are discarded.

The software also undergoes quality control. The engineers testing the software are independent from the engineers developing it. Each test code is checked by two independent testing engineers. Each piece of software is tested extensively against a range of synthetic data (i.e. generated for the purpose of testing) for which the expected output is established independently. If the software fails to return this output, then the software is deemed to have failed the test and is examined and modified accordingly. For statistical software (including that used for propensity score matching, for Kaplan-Meier analysis, etc), an additional quality control step is implemented. Two independent codes are written in two different programming languages (typically R and python) and the statistical results are compared. If discrepancies are identified, then the codes are deemed to have failed the test and are examined and modified accordingly. All the code is reviewed independently by another engineer.

The test strategy follows three levels of granularity:

- 1. Unit tests: These test specific blocks, or units, of code that perform specific actions (e.g. querying the database).
- 2. Integration tests: These ensure that different components are working together correctly.
- 3. End-to-end tests: These tests run the entire system and check the final output.

Some comments on advantages and disadvantages of EHR data

The advantage of EHR data, like those in TriNetX, over insurance claim data is that both insured and uninsured patients are included. An advantage of EHR data over survey data is that they represent the diagnostic rates in the population presenting to healthcare facilities. This provides an accurate account of the burden of specific diagnoses on healthcare systems. The downside of

relying on diagnoses is that they obviously do not account for undiagnosed patients who might be suffering from the illness but did not seek medical attention (or in whom the diagnosis was missed). A general limitation of EHR data is that a patient may be seen in different HCOs for different parts of their care and if one HCO is not part of the federated network then part of their medical records may not be available. Using a network of HCOs (rather than a single HCO) limits this possibility but does not fully remove it. Finally, historical data before the start of EHRs (or the addition of an HCO to the network) may be incomplete.

Cohorts definition and index events

The two control cohorts used consisted of patients who received an mRNA vaccine and patients with a diagnosis of influenza. Specifically, patients who received the vaccine were those who had any of the following procedure codes in their electronic health records:

- 91300: "Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) (Coronavirus disease [COVID-19]) vaccine, mRNA-LNP, spike protein, preservative free, 30 mcg/0.3mL dosage, diluent reconstituted, for intramuscular use"
- 0001A: "Immunization administration by intramuscular injection of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) (Coronavirus disease [COVID-19]) vaccine, mRNA-LNP, spike protein, preservative free, 30 mcg/0.3mL dosage, diluent reconstituted; first dose"
- 0002A: "Immunization administration by intramuscular injection of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) (Coronavirus disease [COVID-19]) vaccine, mRNA-LNP, spike protein, preservative free, 30 mcg/0.3mL dosage, diluent reconstituted; second dose"
- 91301: "Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) (Coronavirus disease [COVID-19]) vaccine, mRNA-LNP, spike protein, preservative free, 100 mcg/0.5mL dosage, for intramuscular use"
- 0011A: "Immunization administration by intramuscular injection of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) (Coronavirus disease [COVID-19]) vaccine, mRNA-LNP, spike protein, preservative free, 100 mcg/0.5mL dosage; first dose"
- 0012A: "Immunization administration by intramuscular injection of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) (Coronavirus disease [COVID-19]) vaccine, mRNA-LNP, spike protein, preservative free, 100 mcg/0.5mL
- 2468231: "SARS-CoV-2 (COVID-19) vaccine, mRNA spike protein"

Patients with influenza were those who had any of the following diagnoses:

- J09: Influenza due to certain identified influenza viruses
- J10: Influenza due to other identified influenza virus
- J11: Influenza due to unidentified influenza virus.

Because some patients with the control index event might have had COVID-19 at a different point in time, we excluded from the control cohorts all those who had COVID-19 at any point in time. To avoid any contamination between cohorts, COVID-19 as an exclusion criterion was defined in the broader sense to be all patients with a confirmed diagnosis of COVID-19 (ICD-10 code U07.1) but also patients with an unconfirmed COVID-19 diagnosis (U07.2), a recorded

positive PCR test for COVID-19, or any of the following recorded on or after January 20, 2020: Pneumonia due to SARS-associated coronavirus (J12.81), Other coronavirus as the cause of disease classified elsewhere (B97.29), or Coronavirus infection unspecified (B34.2). Inclusion of the latter three diagnostic codes captures patients who receive a COVID-19 diagnosis in the early stage of the pandemic when the ICD code for COVID-19 (U07) was not yet defined. Specifically, the following codes were excluded from the control cohorts if they occurred on or after January 20, 2020:

- U07.1: COVID-19, virus identified
- U07.2: COVID-19, virus not identified
- J12.81: Pneumonia due to SARS-associated coronavirus
- B97.29: Other coronavirus as the cause of disease classified elsewhere
- B34.2: Coronavirus infection, unspecified
- Positive SARS-CoV-2 RNA in Respiratory specimen
- Positive SARS-CoV-2 RNA in Unspecified specimen
- Positive SARS-CoV-2 N gene in Respiratory specimen
- Positive SARS-CoV-2 N gene in Unspecified specimen
- Positive SARS-CoV-2 RdRp gene in Respiratory specimen
- Positive SARS-CoV-2 E gene in Respiratory specimen
- Positive SARS-CoV-2 E gene in Unspecified specimen
- Positive SARS-CoV-2 RNA panel in Respiratory specimen
- Positive SARS-CoV-2 RNA panel in Unspecified specimen
- Positive SARS-CoV-2 RNA in Nasopharynx
- Positive SARS coronavirus 2 and related RNA
- Positive SARS-related coronavirus RNA in Respiratory specimen
- Positive SARS coronavirus 2 ORF1ab in Respiratory specimen

Baseline characteristics code

When reporting baseline characteristics, the following ICD-10 codes are used:

- Obesity: E66
- Hypertension: I10-I16
- Chronic kidney disease: N18
- Ischemic heart disease: I20-I25
- Heart failure: I50
- Disease of the arteries, arterioles, or capillaries: I70-I79
- Disease of (non-cerebral) veins: I80-I87
- Cerebral/Pre-cerebral artery stenosis/occlusion: I63 (cerebral infarction), I65 (Occlusion and stenosis of precerebral arteries, not resulting in cerebral infarction), I66 (Occlusion and stenosis of cerebral arteries, not resulting in cerebral infarction)
- Intracranial hemorrhage: I60 (Nontraumatic subarachnoid hemorrhage), I61 (Nontraumatic intracerebral hemorrhage), I62 (Other and unspecified nontraumatic intracranial hemorrhage)
- Dementia: F01 (Vascular dementia), F02 (Dementia in other diseases classified elsewhere), F03 (Unspecified dementia), G30 (Alzheimer's disease), G31.0 (Frontotemporal dementia), and G31.83 (Dementia with Lewy bodies)
- Chronic lower respiratory diseases: J40-J47

Connective tissue disorders: M30-M36

Liver diseases: K70-K77Diabetes mellitus: E08-E13

- Malignancy: C00-C14 (Malignant neoplasms of lip, oral cavity and pharynx), C15-C26 (Malignant neoplasms of digestive organs), C30-C39 (Malignant neoplasms of respiratory and intrathoracic organs), C40-C41 (Malignant neoplasms of bone and articular cartilage), C43-C44 (Melanoma and other malignant neoplasms of skin), C45-C49 (Malignant neoplasms of mesothelial and soft tissue), C50 (Malignant neoplasms of breast), C51-C58 (Malignant neoplasms of female genital organs), C60-C63 (Malignant neoplasms of male genital organs), C64-C68 (Malignant neoplasms of urinary tract), C69-C72 (Malignant neoplasms of eye, brain and other parts of central nervous system), C73-C75 (Malignant neoplasms of thyroid and other endocrine glands), C76-C80 (Malignant neoplasms of ill-defined, other secondary and unspecified sites), C7A (Malignant neoplasms of lymphoid, hematopoietic and related tissue)

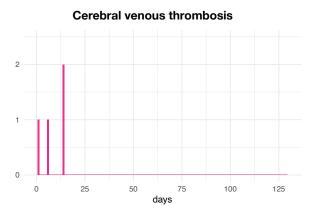
Details of statistical analysis

In propensity score matching, the propensity score was calculated using a logistic regression (implemented by the function LogisticRegression of the scikit-learn package in Python 3.7). To eliminate the influence of ordering of records, the order of the records in the covariate matrix were randomised before matching.

References

Taquet M, Geddes JR, Husain M, *et al.* 6-month neurological and psychiatric outcomes in 236 379 survivors of COVID-19: a retrospective cohort study using electronic health records. *Lancet Psychiatry* Published Online First: 6 April 2021. doi:10.1016/S2215-0366(21)00084-5

Supplementary figures



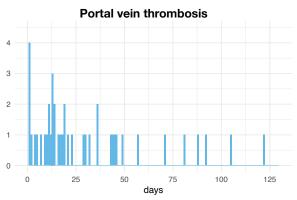


Fig. S1 – Distribution of the day of recorded death relative to the diagnosis of CVT/PVT for patients who died after having had a CVT post COVID-19 (top) or a PVT post COVID-19 (bottom).

Supplementary table

Table S1 – Baseline characteristics of the matched cohorts of patients diagnosed with COVID-19 vs. influenza. SMD=Standardized mean difference.

| | COVID-19 | Influenza | SMD |
|---|---------------|---------------|-------|
| Sample size, n | 392424 | 392424 | - |
| Age, mean (SD), y | 40.9 (20.7) | 41.3 (21.0) | 0.02 |
| Sex, n (%) | | | |
| Female | 218345 (55.6) | 231499 (59.0) | 0.07 |
| Male | 173970 (44.3) | 160872 (41.0) | 0.07 |
| Other | 109 (0.03) | 53 (0.01) | 0.01 |
| Race, n (%) | | | |
| White | 242092 (61.7) | 258548 (65.9) | 0.09 |
| Black or African American | 67373 (17.2) | 65270 (16.6) | 0.01 |
| Asian | 11110 (2.8) | 11573 (2.9) | 0.007 |
| American Indian or Alaska Native | 1653 (0.4) | 1622 (0.4) | 0.001 |
| Native Hawaiian or Other Pacific Islander | 1083 (0.3) | 1291 (0.3) | 0.01 |
| Unknown | 69113 (17.6) | 54120 (13.8) | 0.1 |

Table S2 – Baseline characteristics of the matched cohorts of patients diagnosed with COVID-19 and people receiving an mRNA vaccine. SMD=Standardized mean difference.

| | COVID-19 | mRNA Vaccine | SMD |
|---|---------------|---------------|----------|
| Sample size, n | 366869 | 366869 | - |
| Age, mean (SD), y | 55.2 (18.1) | 55.0 (18.1) | 0.01 |
| Sex , n (%) | | | |
| Female | 207968 (56.7) | 210226 (57.3) | 0.01 |
| Male | 158734 (43.3) | 156536 (42.7) | 0.01 |
| Other | 167 (0.05) | 107 (0.03) | 0.008 |
| Race, n (%) | | | |
| White | 248653 (67.8) | 245477 (66.9) | 0.02 |
| Black or African American | 52633 (14.3) | 52069 (14.2) | 0.004 |
| Asian | 13385 (3.6) | 16959 (4.6) | 0.05 |
| American Indian or Alaska Native | 1878 (0.5) | 2042 (0.6) | 0.006 |
| Native Hawaiian or Other Pacific Islander | 718 (0.2) | 746 (0.2) | 0.002 |
| Unknown | 49602 (13.5) | 49576 (13.5) | 2.00E-04 |

STROBE Statement—Checklist of items that should be included in reports of *cohort studies*

| | Item | |
|-------------------------|----------|---|
| TRY 1 1 1 4 | No | Recommendation |
| Title and abstract | 1 | (a) Indicate the study's design with a commonly used term in the title or the abstract |
| | | (b) Provide in the abstract an informative and balanced summary of what was done and what was found |
| Introduction | | |
| Background/rationale | 2 | Explain the scientific background and rationale for the investigation being reported |
| Objectives | 3 | State specific objectives, including any prespecified hypotheses |
| Methods | | |
| Study design | 4 | Present key elements of study design early in the paper |
| Setting | 5 | Describe the setting, locations, and relevant dates, including periods of recruitment, exposure, follow-up, and data collection |
| Participants | 6 | (a) Give the eligibility criteria, and the sources and methods of selection of |
| 1 41 41 41 41 41 | Ü | participants. Describe methods of follow-up |
| | | (b) For matched studies, give matching criteria and number of exposed and unexposed |
| Variables | 7 | Clearly define all outcomes, exposures, predictors, potential confounders, and |
| | | effect modifiers. Give diagnostic criteria, if applicable |
| Data sources/ | 8* | For each variable of interest, give sources of data and details of methods of |
| measurement | | assessment (measurement). Describe comparability of assessment methods if |
| | | there is more than one group |
| Bias | 9 | Describe any efforts to address potential sources of bias |
| Study size | 10 | Explain how the study size was arrived at |
| Quantitative variables | 11 | Explain how quantitative variables were handled in the analyses. If applicable, describe which groupings were chosen and why |
| Statistical methods | 12 | (a) Describe all statistical methods, including those used to control for |
| | | confounding |
| | | (b) Describe any methods used to examine subgroups and interactions |
| | | (c) Explain how missing data were addressed |
| | | (d) If applicable, explain how loss to follow-up was addressed |
| | | (\underline{e}) Describe any sensitivity analyses |
| Results | | |
| Participants | 13* | (a) Report numbers of individuals at each stage of study—eg numbers |
| | | potentially eligible, examined for eligibility, confirmed eligible, included in |
| | | the study, completing follow-up, and analysed |
| | | (b) Give reasons for non-participation at each stage |
| | | (c) Consider use of a flow diagram |
| Descriptive data | 14* | (a) Give characteristics of study participants (eg demographic, clinical, social) |
| | | and information on exposures and potential confounders |
| | | (b) Indicate number of participants with missing data for each variable of |
| | | interest |
| | . | (c) Summarise follow-up time (eg, average and total amount) |
| Outcome data | 15* | Report numbers of outcome events or summary measures over time |
| Main results | 16 | (a) Give unadjusted estimates and, if applicable, confounder-adjusted |
| | | estimates and their precision (eg, 95% confidence interval). Make clear which |
| | | confounders were adjusted for and why they were included |
| | | (b) Report category boundaries when continuous variables were categorized |
| | | (c) If relevant, consider translating estimates of relative risk into absolute risk |
| | | for a meaningful time period |

| Other analyses | 17 | Report other analyses done—eg analyses of subgroups and interactions, and sensitivity analyses |
|-------------------|----|--|
| Discussion | | |
| Key results | 18 | Summarise key results with reference to study objectives |
| Limitations | 19 | Discuss limitations of the study, taking into account sources of potential bias or imprecision. Discuss both direction and magnitude of any potential bias |
| Interpretation | 20 | Give a cautious overall interpretation of results considering objectives, limitations, multiplicity of analyses, results from similar studies, and other relevant evidence |
| Generalisability | 21 | Discuss the generalisability (external validity) of the study results |
| Other information | | |
| Funding | 22 | Give the source of funding and the role of the funders for the present study and, if applicable, for the original study on which the present article is based |

^{*}Give information separately for exposed and unexposed groups.

Note: An Explanation and Elaboration article discusses each checklist item and gives methodological background and published examples of transparent reporting. The STROBE checklist is best used in conjunction with this article (freely available on the Web sites of PLoS Medicine at http://www.plosmedicine.org/, Annals of Internal Medicine at http://www.annals.org/, and Epidemiology at http://www.epidem.com/). Information on the STROBE Initiative is available at http://www.strobe-statement.org.